

Amendments to the Specification:

Please replace the paragraph beginning at page 39, line 11, with the following amended paragraph:

AcY_sEY_sLDY_sDF (SEQ ID NO 2): The peptide was synthesized on 0.025 mmol scale by standard coupling procedures. Commercially available pre-loaded resin (0.5 mmol/g) was used. The N-terminal Fmoc group was cleaved and the amino group acetylated. The azidomethylene group was cleaved in the usual way. Sulfation was performed as described above. Cleavage and lyophilization affords 28 mg of a white solid. HPLC purification [Alltech Econosil C18, one major peak: $t_2 = 22.76$ min, gradient system: CH₃CN/0.1 M aq. NH₄OAc 5% – 75% in 40 min, 8 mL/min] affords 8 mg (27% based on resin loading, minus resin for characterization) of a flocculent white solid. IR (KBr): 1244 br, str, 1050 br, str; LRMS (MALDI, α -cyano-4- hydroxycinnamic acid matrix, negative ion mode): m/z 1138.3 [calc'd M-3SO₄+NH₄⁺ 1138.48].

Please replace the paragraph beginning at page 39, line 21 with the following amended paragraph:

AcYEYLDYDF (SEQ ID NO 4): A fraction of the phenol-protected material (10 mg resin) from the synthesis of AcY_sEY_sLDY_sDF (SEQ ID NO 2) above was cleaved (yield 6 mg), dissolved in MeOH/H₂O (2 mL) and subjected to hydrogenation over Pearlman's catalyst (10 mg) for 12h under an H₂ filled balloon. Filtration through pre-rinsed Celite (MeOH/H₂O, 1:1 eluant) afforded 3 mg crude material after lyophilization. LRMS (FAB α -cyano-4-hydroxycinnamic acid matrix, positive ion mode): m/z 1215.4 [calc'd MH+ 2Na⁺ 1215.44].

Please replace the paragraph beginning at page 39, line 29, with the following amended paragraph:

AcYEY_sLDYDF (SEQ ID NO 3): The solid phase synthesis was performed according to the general procedures described above. Cleavage from the resin gave 17 mg of crude peptide. This material was subjected to hydrogenation over Pearlman's catalyst (20 mg) for 12h under an H₂ filled balloon. Filtration through pre-rinsed Celite (H₂O eluant). This material was subjected to HPLC (Alltech Econosil C18) gave three major peaks, two of which appeared to be deletion peptides (by MALDI-MS, we were unable to assign a structure based on the mass spectra, however the peptides appeared to be sulfated, as judged by HPLC retention time). The longest retained peptide (*t_r* = 33.48 min, gradient system: CH₃CN/ 0.1 M aq. NH₄OAc 5% – 75% in 40 min, 8 mL/min] pooling of this HPLC fraction and lyophilization afforded the desired peptide as a fluffy white solid (4.6 mg, 5.2 %) LRMS (MALDI, α -cyano-4-hydroxycinnamic acid matrix, negative ion mode): *m/z* 1170.4 [calc'd M-SO₄⁻+NH₄⁺ 1169.42]; (MALDI, 2,4,6- trihydroxyaceto-phenone, negative ion mode): *m/z* 1191.6 [calc'd M – SO₃ + Na⁺ for C₅₇H₆₈N₈O₂₂S 1191.41] IR (KBr): 1256 br, str, 1049 br, str.

Please replace the paragraph beginning at page 43, line 3, with the following amended paragraph:

pEY_sLDYDF (SEQ ID NO 5): This peptide was generated in the attempted synthesis of Fmoc Y_sEY_sLDY_sDF (SEQ ID NO 2) *via* the stepwise protocol described above. HPLC purification of the product and analytical characterization revealed the pyroglutamate-terminated structure. A satisfactory mass spectrum was not obtained for this compound. However, 2-dimensional ¹HNMR analysis (TOCSY, COSY) showed, unambiguously, this sequence. HPLC [Econosil C18, gradient 25:75 CH₃CN/0.1M aqueous NH₄OAc – 75:25/30 – 40min at 8 mL/min; retention time: 36.8 min].